

General Laboratory Considerations & Recommendations for *Candida auris* (*C. auris*) Identification

All yeast specimen obtained from sterile sites (e.g., bloodstream) should be identified to the species level.

Identification of yeast and/or *Candida* presence in non-sterile body sites (e.g., urine, sputum, skin), may also represent colonization and/or infection, and speciation should be considered.

All facilities should consider germ tube testing when yeast is isolated from non-sterile sites to differentiate *Candida albicans* from other *Candida* species as a first step to determine the necessity for speciation. If germ tube testing is negative, further speciation is indicated.

Consider expanding identification of any yeast isolated on routine bacterial or fungal cultures from patients who have multiple risk factors for *C. auris* colonization and infection.

Facility-based admission screening should be considered for high-risk patients and for all patients newly admitted to facilities with suspected and/or documented *C. auris* transmission.

C. auris can be misidentified as other *Candida* species with certain laboratory instruments.

Ensure that your laboratory methods can accurately identify *C. auris*.

C. auris Testing Efforts Typically Supported by NJDOH

1. Confirmatory testing via MALDI-TOF
2. Antimicrobial susceptibility testing
3. Surveillance support for healthcare facilities with documented or suspected transmission

Identification Method	Organisms <i>C. auris</i> can be misidentified as
Vitek 2 YST	<i>Candida haemulonii</i> <i>Candida duobushaemulonii</i>
API 20C	<i>Rhodotorula glutinis</i> (characteristic red color not present) <i>Candida sake</i>
API ID 32C	<i>Candida intermedia</i> <i>Candida sake</i> <i>Saccharomyces kluyveri</i>
BD Phoenix yeast identification system	<i>Candida haemulonii</i> <i>Candida catenulata</i>
MicroScan	<i>Candida famata</i> <i>Candida guilliermondii</i> <i>Candida lusitaniae</i> <i>Candida parapsilosis</i>
RapID Yeast Plus	<i>Candida parapsilosis</i>

Resources for Implementing Facility-Based Admission or Surveillance Screening

Although some commercial laboratories have not yet implemented molecular assays (e.g., PCR) for surveillance or screening swabs, major commercial laboratories have validated gold-standard, MALDI-TOF capabilities and can perform fungal culture on surveillance/screening swabs.

Commercial Laboratory	Identification Method	Test (Order Code)
ARUP	Fungal culture (must indicate "rule out <i>Candida auris</i> ")	<ul style="list-style-type: none">Fungal culture, yeast (0060149)Yeast ID – MALDI Bruker; sequencing if no ID (0060163)
LabCorp	Fungal culture (must indicate "rule out <i>Candida auris</i> ")	<ul style="list-style-type: none">Fungal culture, yeast (008482)
Mayo	PCR	<ul style="list-style-type: none"><i>Candida auris</i> surveillance (PCR) (CAURS 607883)Yeast ID – MALDI Bruker; sequencing if no ID (FUNID 8223)
Quest	Fungal culture (must indicate "rule out <i>Candida auris</i> ")	<ul style="list-style-type: none">Fungal culture, yeast (20541)Yeast ID – MALDI Vitek MS or Bruker; sequencing if no ID (39507)

For clinical laboratories interested in establishing and validating their own methods for *C. auris* identification, please find CDC's recommended protocols for processing screening swabs by scanning here!

